

CHRONIC MASTITIS: LEADING CAUSE OF UDDER FIBROSIS AND DIFFERENT MEANS OF ITS MANAGEMENT

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ABSTRACT

Bovine mastitis is an inflammatory response mainly caused by bacteria with substantial economic losses to the world dairy industry and brings issues like food safety for humans. The long-term presence of inflammation of mammary tissue causes chronic mastitis that leads to udder fibrosis. Udder fibrosis is a major problem for dairy cattle, which drops the milk yield. Mastitis at an early stage of lactation results in long-term production loss. In contrast, subclinical mastitis causes an increased risk of getting clinical mastitis in subsequent lactation and premature culling of animals. Risk factors including husbandry practices, milking methods, temperature fluctuation, and irrational use of antibiotics are significantly associated with mastitis. Consequently, the early detection of mastitis is crucial, considering the difficulty of detecting sub-clinical mastitis. So, we can use diagnostic tools such as different chemical tests, ultrasonography, teat endoscopy, and physical examination of the fibrosed udder. It is possible to treat mastitis in early-stage; however, it becomes difficult to treat once the fibrosis occurs. The current manuscript covers a detailed review of published data on mastitis, udder fibrosis causes, manageable factors, and other issues that merit further investigation.

Keywords: Mastitis, Subclinical mastitis, Risk Factors, Udder fibrosis, Mastitis management

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1. INTRODUCTION

The most transmissible disease of dairy animals, which affects the quality and quantity of milk produced globally, is mastitis (Balkwill 2004). Mastitis is a dangerous disease of dairy animals and causes massive economic losses (Pal et al. 2019; Du et al. 2022) and occurs repeatedly (Halasa et al. 2007). Chronically infected quarters of the mammary tissue cause lower milk yield (Gonçalves et al. 2020). Mastitis prevalence rate in cow estimate falls within the range of 23.2 and 81.1% for the country (Girma et al. 2012; Zenebe et al. 2014). According to an estimate, an affected quarter suffers 30% reduction in productivity and an affected cow loses 15% of its production for the lactation (Radostits et al. 2007). The economic implication of bovine mastitis is derived from the high costs of diagnosis, treatment, loss of milk production, early culling and cost of the control program in subclinical and

clinical cases (Khan et al. 2013; Hussain et al. 2013; Qayyum et al. 2016a, 2016b; Ji et al. 2020). Subsequently, the disease has been recognized as one of the major constraints of the dairy sector that needs attention. In recent studies, the work is being done to find the role of the fibroblasts (which were collected from the mastitis infected tissue of the udder of the cow) in the increase of the inflammation and the fibrosis of the mammary tissue (He et al. 2017).

In mastitis, the tissue fibrosis occurred due to too much production of extracellular matrix (ECM) (Zou et al. 2017). Abnormally proliferated fibroblasts, which are activated during mastitis, replace the damaged tissue and produce excessive ECM (Ghosh et al. 2013). Epithelial cells can produce fibroblasts and myofibroblasts by a process named Epithelial-mesenchymal transition (EMT) (Canisso et al. 2021). Epithelial cells undergo the transition to a mesenchymal phenotype to produce these fibroblasts and myofibroblasts (Zou et al. 2017). The main stromal cells of the mammy glands of the bovines are mammary fibroblasts. These fibroblasts control the epithelial cell behavior by cell-to-cell interaction, these interactions may be direct or indirect. Stromal fibroblasts also lead to persistent inflammation of the mammary gland by secreting cytokines, chemokines and growth factors (Buckley 2011; Hasan et al. 2022). Fibroblasts are the important role in changing acute inflammation to adaptive immunity and tissue repair because fibroblasts are the important soldier cells in the immune system (Buckley et al. 2001). The fibroblasts of the diseased tissue show a different type of composition compared with fibroblasts taken from the normal tissue at the same location from the body (Hogaboam et al. 1998; Xu et al. 2007).

Inflammation can stay for a long time when the immune system is overstimulated. The fibroblasts have a long life, and they are present in the inflammation abundantly. Due to their longer life, they can stay in the site of the inflammation for a longer time and may lead to hyperstimulation of the immune system. A control mechanism should be present to avoid the overstimulation of the immune system (Brouty-Boyé et al. 2000; Xu et al. 2007). The molecular basis for the persistently present, activated fibroblasts at sites of chronic inflammation is not yet understood (Buckley et al. 2001). Fig. 1 shows the inflammation of the udder indicating chronic case of mastitis that may leads to the udder fibrosis.



Fig. 1: Fibrosed udder after chronic mastitis (left hind quarter teat).



Fig. 2: Flow Diagram showing etiology of Bovine Mastitis involving different parameters.

1.1. Etiology

The main causative pathogens of the mastitis vary as reported in various studies (Cobirka et al. 2020). Most important pathogen accountable for mastitis in dairy animals is the coagulase-positive Staphylococcus (S.) aureus. Coagulase-negative Staphylococci can also cause clinical or subclinical mastitis (Semik-Gurgul et al. 2021). Even the Gram-positive pathogens are responsible for the severe cases of the mastitis in dairy (Schmenger and Krömker 2020). The 90% of the Staphylococcus species strains isolated from the mastitis-infected animals were S. aureus (Freitas et al. 2018; Du et al. 2022).

Mastitis is caused by a number of the viruses like (Infectious rhinotracheitis, vesicular stomatitis), Bacteria include (S. aureus, Streptococcus (St.) agalactia, St. dysgalactiae, St. faecalis, St. pyogens, Corynebacterium pyogens, St. zooepidemicus, Klebsiella species, Mycobacterium bovis, Escherichia coli, Brucella abortus, Pseudomonas pyocyaneus, Pasteurella maltocida, Leptospira Pomona, Mycoplasma bovis, and Mycoplasma bovigenitalium), Molds (Aspergillus fumigatus), rickettsia, and yeast (Sarma and Hussain 2021).

These all may cause mastitis in different forms i.e. subclinical mastitis caused by the *S. aureus* and the Streptococcus species. Clinical mastitis and per acute mastitis are commonly caused by the *Pseudomonas aeruginosa, Escherichia coli, S. aureus*, and *St. dysgalactiae*. Acute mastitis is commonly caused by coliform organisms, Subacute mastitis, and chronic mastitis caused by the *St. uberis*, and *S. aureus*. Chronic mastitis lasts long and leads towards the fibrosis of the udder and the affected part of the udder becomes hard which can ultimately lead to the loss of the function of the udder (Sarma and Hussain 2021).

1.2. Pathogenesis

S. aureus in a persistent infection can cause the chronic form of the mastitis by escaping through macrophage phagocytosis and produces mammary gland fibrosis (Bi et al. 2020). The process of chronic mastitis infection is that the mastitis causing bacteria can live in the host phagocytes and some non-phagocytic cells as well as mammary epithelial cells without being engulfed or phagocytosed; in these cells the antibiotics cannot reach up to the proper concentration failing to trigger innate and acquired immune responses properly. The exact mechanism of the immune responses in the memory tissue is not yet known. There may be chances of immune suppression (Wang et al. 2015).

In mastitis pathogenesis, various cells can generate an immune response like neutrophils (Sladek et al. 2005) macrophages (Sladek et al. 2006) lymphocytes (Sordillo and Streicher 2002) and epithelial cells in the mammary gland (Schukken et al. 2011). A cytokine from the chemokine group is known as a chemoattractant for the lymphocytes called stromal cell-derived factor1(SDF-1), also known as CXCL12 (Kobayashi et al. 2014). In chronic inflammation (as in case of the chronic mastitis), lymphocytes and the dendritic cells are accumulated and stay for a long time at the site of the inflammation because the fibroblasts which are producing the SDF-1, started the overproduction of the SDF-1, which is the chemoattractant for these lymphocytes and the dendritic cells. The SDF-1 (which is secreted by the cancerous fibroblasts) is attached to the receptors (CXCR7 and CXCR4) of these cancerous cells surfaces, and they started to downstream the intracellular signal pathways that control the metastasis, angiogenesis and drug-resistance of these cancer cells (Hattermann et al. 2014). SDF-1 enhanced the breast cancer cells (Kang et al. 2005). SDF-1 also initiates the migration, invasion, survival, proliferation, and adhesion in the cell (Balkwill 2004; Luker and Luker 2006). Even though SDF-1 performs these functions in the cell but still little is known about its role in the start of the EMT and the production of the inflammation in the mammary cells of the bovines.

An experiment found that inflammation-associated fibroblasts (INFs) expressed elevated amounts of vimentin and collagen-1 compared to the normal fibroblasts (NFs). Vimentin is involved in the rigidity of the cells. The skeleton of the collagen-1 can supports the accumulation of the cells like the fibroblast (Oliveira et al. 2010), which in turn produces more and more ECM and other types of the proteins responsible for the fibrosis. Still, INFs expressed a lesser Matrix metalloproteinase-1 (MMP-1) level than NFs. The MMP-1 is involved in the breakdown of the ECM. This MMP-1 is less produced by the INFs and contributes to an increase in the ECM, which can cause fibrosis. Fibroblasts can also secrete cytokines, chemokines and growth factors which can contribute to the persistent inflammation of the mammary glands (Buckley et al. 2001; Shah et al. 2022).

1.3. Predisposing Factors to Mastitis Leading Towards Udder Fibrosis

Chilling increases the inflammation of the udder, which is already infected, but this is not proved experimentally. These factors include the feed, age, stage of lactation, teat characteristics, hand and machine milking, and vacuum level (Belay et al. 2022). The rate of udder infection increases with age (Plastridge 1958). Some unknown changes associated with age may be related to susceptibility to St. agalactiae infection (Mvrphy 1947; Lancaster and Stuart 1949). Stage of the lactation also matters, but St. agalactiae infection is not related to the stage of lactation (Plastige et al. 1942). Injuries to teats by stepping the cows to the teat, cuts, lesions, surgeries, can result in the entry of the many kinds of microorganisms inside the mammary tissue, cause inflammation and further aggravate the teat infection (Plastridge 1958). This persistent inflammation can thus lead to fibrosis due to the persistent infection in the mammary tissue (Wang et al. 2015). It is stated earlier that there is an increased prevalence of mastitis in machine-milked animals than in animals milked with using hand. In the latest instances, it is observed that the palms of milker may be a primary source in the spread of udder infections. A vacuum level of over 15 inches of mercury (pressure used in the milking machine) be likely to increase the occurrence of mastitis



(Burkey and Sanders 1949). Fig. 3 depicted various factors associated with bovine mastitis and development of fibrosis in the udder.

1.4. Risk Factors

There are many causes of udder fibrosis, but our main concern is mastitis and inflammation related to inflammation. Many factors are responsible for the induction of clinical mastitis i.e., Herd (Schukken et al. 1991; Nyman et al. 2007; Belay et al. 2022) and the factors associated with cow (Barkema et al. 1998; Suriyasathaporn et al. 2000; Riekerink et al. 2007). Among factors associated with cow, somatic cell count is the most important factor in assessing clinical mastitis, with the very low somatic cell count (Suriyasathaporn et al. 2000) and the high somatic cell count (Beaudeau et al. 1998). Heifers have a very low number of clinical mastitis cases except for the



Fig. 3: Factors associated with bovine mastitis and udder fibrosis.



Fig 4: Affected teats showing swelling, heat, hardness, redness, watery appearance, flakes and pus clots in the milk.

first week of lactation (Barkema et al. 1998; Mohanty et al. 2019). A cow that has clinical mastitis previously will be more prone to clinical mastitis during the next lactation (Houben et al. 1993; Zadoks et al. 2001). Clinical mastitis mostly occurs during the starting days of lactation (Miltenburg et al. 1996). The relationship between the cow-related factors and the pathogen-associated clinical mastitis is not often studied (Zadoks et al. 2001; De Haas et al. 2002; De Haas et al. 2004).

In dairy cattle, genetic variation (for mastitis resistance) is present, generally accepted. The somatic cell count and the clinical causes are the phenotypic traits of mastitis. A genetic correlation exists among these traits; many studies are being carried out to study this type of polygenic variation of the traits. From the economic point of view, consumer concern, welfare, and food safety, it is necessary to add mastitis to the breeding objective of the dairy cattle breeds. Improvements in the selection of the mastitis resistance genetics of the dairy cattle are being studied, and it includes advances in the modeling and udder morphology (Rupp and Boichard 2003).

1.5. Diagnosis

Clinical observations or direct or indirect measures of the inflammatory response to infection are the base of the diagnosis of mastitis (Adkins and Middleton 2018). Clinical mastitis may be noticed with the aid of modifications in the appearance of milk, swelling, redness, and elevation in udder temperature. However, animals with subclinical mastitis may be detected most effectively through laboratory tests as subclinical mastitis does not display any gross changes in milk or udder (Reza et al. 2011; Mohanty et al. 2019). Early diagnosis of mastitis is vital because changes in the udder tissue occur much earlier before they become apparent. The prognosis of subclinical mastitis is difficult to interpret as milk seems to be normal physically (Canisso et al. 2021). Numerous strategies are used to diagnose subclinical mastitis, primarily based totally on physical and chemical modifications of milk and isolation of organisms (Batra and McAllister 1984; Emanuelson et al. 1987). The present study indicates that the surf field mastitis test is a very sensitive test that detected the highest number of subclinical mastitis (46%) and clinical mastitis (8%) (Kurjogi and Kaliwal 2014).



Physical examination of the mammary tissue and the teats is carried out to touch the mammary tissue externally. The consistency is felt by the touch sense., If the consistency feels hard it means there may be fibrosis present. Secondly, there may be the blockage of the teat and the milk that can affect the milk let down, as there are four quarters of the udder and every quarter has a teat attach to it, so the fibrosis may take place in quarter of the udder or may be to all four quarters of the udder. The blockage due to fibrosis may be partial or complete (Pal et al. 2019). Fig. 4 shows affected teat with signs of swelling, heat, hardness, redness, or pain to animals as it was reluctant to touch. That animal's milk had watery appearance, flakes, and clots with pus.

Ultrasonography is used to examine the internal structures of the mammary tissue by using non-invasive techniques. The images are obtained, and thus the internal structures are examined thoroughly, and the type of the injury is determined (Franz et al. 2009). There is another device named, Theloscopy (Teat endoscopy) that is used to visualize the internal structures of the udder and the teat; this technique is very reliable to determine the problems related to the blockage of the teat (Canisso et al. 2021). By this technique, the severity and the degree of the injury to the mammary tissue can be determined (Rathod et al. 2009).

1.6. Treatment

Mastitis can lead to fibrosis when not treated. The fibrotic tissue can further proliferate and take the place of the remaining normal soft tissue of the mammary gland (Umadevi et al. 2015). In an experiment, the fibrosed mammary tissue cows were treated with Pendistrin SH intramammary infusion 24 hours for one week and topical massage of Mastilep gel twice a day for 10-15 days. A healing rate of 93% was observed in all the treated cows between 10-15 days (Umadevi et al. 2015).

The formation of fibrous tissue and encircling the pathogen is a shielding mechanism from stopping the propagation of the pathogen. Thus, it considerably hardens the udder and teat cisternae (Nieberle and Cohrs 1966). When fibrous tissue proliferates, either consecutive abscess formation or spontaneous convalescence occurs and literature confirmed no treatment regimen. The infected cow is restrained properly, then fibrosed material from the fibrosed cow udder, teat is crushed by the teat bistoury, and the hand milking removes this material. Then to avoid further adhesion in the teat canals, four plastic tubes are passed through the teat canal (Radostits et al. 2007) followed by the insertion of the intra-mammary tubes for three consecutive days. Antibiotics and anti-inflammatory drugs are also administered. After this treatment, animal shows signs of recovery as the milk comes out of the teat canal both in the amount and quality (Ijaz et al. 2014). The intramammary injection of the Bifidobacterium breve can remove the minor pathogens associated with the mastitis and lower the somatic cell count in the affected quarters of the udder of the dairy cattle. It is the non-antibiotic method to cure chronic subclinical mastitis (Nagahata et al. 2020).

In cases of persistent mastitis, infected mastitis, and neoplastic or hyperplastic conditions of the udder, the radical mastectomy (unilateral or bilateral) is revealed as a recovery process (Andreasen et al. 1993; El-Maghraby 2001; Cable et al. 2004). In many different conditions and persistent suppurative mastitis, pendulous udder, persistent obstructive mastitis, and irreversible udder injuries, mastectomy is specified to remedy inflamed mastitis (Hofmeyr and Oehme 1988). The hemorrhagic complications during a partial mastectomy are very severe because the impacted udder increases in size and is extremely vascularized (Youssef 1999). In affiliation with vascular ligation for mastectomy in cattle, exclusion of the impacted teat has also been described. In cases of chronic mastitis, mastectomy is usually recommended as a pain-relieving technique for enormous lesions involving udder and teat (Cable et al. 2004).

1.7. Homeopathic Treatment of Fibrosed Udder

The fibrotic mastitis cows were treated by oral homeopathic remedies like Silicea 200c and Calcarica flour 200c for 20 days, with a recovery rate of 46.45% (Shah et al. 2010). Homeopathic remedies Phellandrium 30c and Carboanimalis 30c were given orally for 21 days to the various cows having differing degrees of udder fibrosis. The recovery rate was 64.28% (Makkar 2017). The clinically chronic fibrosed mastitis cows were treated with the highest recovery rate of 89% by the homeopathic remedies (which are given orally) like Carboanimalis 200c, Silicea 200c and Conium maculatum 200c for 20 days with most of the cows revered in 14.5 days to 27 days (Karthick 2020).

1.8. Control and Prevention

Early researchers believe that mastitis management was centered on stopping new infections in healthy cows and decreasing the period of infection (Ruegg 2017). A vaccination schedule should be followed to prevent mastitis in dairy cows. Vaccination should be done with the S. aureus bacterins. Wash the teat and udder with warm water and disinfectant. Dipping the teats before and after the milking helps to control the mastitis. Hygiene should be followed; therefore, wear gloves before the milking. In milking machine farms, the machines should be maintained



regularly; the vacuum level should be maintained. Cow dry procedures should be followed strictly. The bedding material should be bacteria-free to control the spread of the infection (Sarma and Hussain 2021). To effectively manage chronic clinical mastitis, preventive methods to avoid new infections and the advancement in treatment programs are required (Wente et al. 2020). A very annoying part of clinical mastitis is the recurrent infection (Jamali et al. 2018).

Separate the infected cows from the healthy cows. To prevent the formation of the fibrosis of the mammary tissue due to chronic mastitis, early detection of mastitis is very important (Mimoune et al. 2021). Early detection of mastitis is always helpful for effective control by timely culling the heavily infected cows.

Conclusion: Mastitis records high economic losses in the dairy industry. This may be a temporary loss in terms of the decrease in milk production and later recovered from the mastitis and animal's parameters get back to the normal regarding milk. Significant changes were observed in the quality and quantity of the milk. As the pathogenic microorganisms enter the udder, they multiply and deteriorate the anatomy and physiology of the udder. It may lead to a permanent loss in the form of udder fibrosis if the chronic phase persists for a longer time. To avoid chronic clinical mastitis, routine tests for subclinical mastitis detection should be done. By controlling the risk factors and proper pasture management strategies, the ratio of diseased animals can be minimized. Observing the pathway of mastitis pathogenesis and transmission methods, better diagnostic tools can be adopted to lessen the burden of mastitis from the dairy industry.

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REFERENCES

Adkins PR and Middleton JR, 2018. Methods for diagnosing mastitis. Veterinary Clinics: Food Animal Practice 34: 479-491. https://doi.org/10.1016/j.cvfa.2018.07.003

- Andreasen CB, Huber MJ and Mattoon JS, 1993. Unilateral fibroepithelial hyperplasia of the mammary gland in a goat. Journal of the American Veterinary Medical Association 202: 1279-1280.
- Balkwill F, 2004. Cancer and the chemokine network. Nature Reviews Cancer 4: 540-550. https://doi.org/10.1038/nrc1388
- Barkema HW, Schukken YH, Lam TJGM, Beiboer ML, Wilmink H, Benedictus G and Brand A, 1998. Incidence of clinical mastitis in dairy herds grouped in three categories by bulk milk somatic cell counts. Journal of Dairy Science 81: 411-419. <u>https://doi.org/10.3168/jds.S0022-0302(98)75591-2</u>
- Batra TR and McAllister AJ, 1984. "A comparison of mastitis detection methods in dairy cattle," Canadian Journal of Animal Science 64: 305–312. <u>https://doi.org/10.4141/cjas84-036</u>
- Beaudeau F, Seegers H, Fourichon C and Hortet P, 1998. Association between milk somatic cell counts up to 400,000 cells/ml and clinical mastitis in French Holstein cows. Veterinary Record 143: 685-687. <u>https://doi.org/10.1136/vr.143.25.685</u>
- Belay N, Mohammed N and Seyoum W, 2022. Bovine mastitis: Prevalence, risk factors, and bacterial pathogens isolated in lactating cows in Gamo Zone, Southern Ethiopia. Veterinary Medicine: Research and Reports 13: 9. <u>https://doi.org/10.2147/VMRR.S344024</u>
- Bi Y, Ding Y, Wu J, Miao Z, Wang J and Wang F, 2020. *Staphylococcus aureus* induces mammary gland fibrosis through activating the TLR/NF-κB and TLR/AP-1 signaling pathways in mice. Microbial Pathogenesis 148: 104427. https://doi.org/10.1016/j.micpath.2020.104427



- Brouty-Boyé D, Pottin-Clémenceau C, Doucet C, Jasmin C and Azzarone B, 2000. Chemokines and CD40 expression in human fibroblasts. European Journal of Immunology 30: 914-919. <u>https://doi.org/10.1002/1521-4141(200003)30:3<914::AID-IMMU914>3.0.CO;2-D</u>
- Buckley CD, 2011. Why does chronic inflammation persist: An unexpected role for fibroblasts. Immunology Letters 138: 12-14. https://doi.org/10.1016/j.imlet.2011.02.010
- Buckley CD, Pilling D, Lord JM, Akbar AN, Scheel-Toellner D and Salmon M, 2001. Fibroblasts regulate the switch from acute resolving to chronic persistent inflammation. Trends in Immunology 22: 199–204. <u>https://doi.org/10.1016/S1471-4906(01)01863-4</u>
- Burkey LA and Sanders GP, 1949. The Significance of Machine Milking in the Etiology and Spread of Bovine Mastitis: A Review. US Department of Agriculture, Bureau of Dairy Industry.
- Cable CS, Peery K and Fubini SL, 2004. Radical mastectomy in 20 ruminants. Veterinary Surgery 33: 263-266. https://doi.org/10.1111/j.1532-950X.2004.04038.x
- Canisso IF, Podico G and Ellerbrock RE, 2021. Diagnosis and treatment of mastitis in mares. Equine Veterinary Education 33: 320-326. https://doi.org/10.1111/eve.13228
- Cobirka M, Tancin V and Slama P, 2020. Epidemiology and classification of mastitis. Animals 10: 2212. https://doi.org/10.3390/ani10122212
- De Haas Y, Barkema HW and Veerkamp RF, 2002. The effect of pathogen-specific clinical mastitis on the lactation curve for somatic cell count. Journal of Dairy Science 85: 1314-1323. <u>https://doi.org/10.3168/jds.S0022-0302(02)74196-9</u>
- De Haas Y, Veerkamp RF, Barkema HW, Gröhn YT and Schukken YH, 2004. Associations between pathogen-specific cases of clinical mastitis and somatic cell count patterns. Journal of Dairy Science 87: 95-105. <u>https://doi.org/10.3168/jds.S00220-302(04)73146-X</u>
- Du XX, Sherein SA, Liu P, Haque MA and Khan A, 2022. Bovine mastitis: Behavioral changes, treatment and control. Continental Veterinary Journal.
- El-Maghraby HM, 2001. Comparison of two surgical techniques for mastectomy of goats. Small Ruminant Research 40: 215-221. https://doi.org/10.1016/S0921-4488(01)00173-0
- Emanuelson U, Olsson T and Holmberg O, 1987. "Comparison of some screening tests for detecting mastitis," Journal of Dairy Science 70: 880–887. <u>https://doi.org/10.3168/jds.S0022-0302(87)80087-5</u>
- Franz S, Floek M and Hofmann-Parisot M, 2009. Ultrasonography of the bovine udder and teat. The Veterinary clinics of North America. Food Animal Practice 25: 669-85. DOI: <u>https://doi.org/10.1016/j.cvfa.2009.07.007</u>
- Freitas CH, Mendes JF, Villarreal PV, Santos PR, Gonçalves CL, Gonzales HL and Nascente PS, 2018. Identification and antimicrobial susceptibility profile of bacteria causing bovine mastitis from dairy farms in Pelotas, Rio Grande do Sul. Brazilian Journal of Biology 78: 661-666. <u>https://doi.org/10.1590/1519-6984.170727</u>
- Ghosh AK, Quaggin SE and Vaughan DE, 2013. Molecular basis of organ fibrosis: potential therapeutic approaches. Experimental Biology and Medicine 238: 461-481. <u>https://doi.org/10.1177/1535370213489441</u>
- Girma S, Mammo A, Bogele K, Sori T, Tadesse F and Jibat T, 2012. Study on prevalence of bovine mastitis and its major causative agents in West Harerghe zone, Doba district, Ethiopia. Journal of Veterinary Medicine and Animal Health 4: 116–123. <u>https://doi.org/10.5897/JVMAH</u>
- Gonçalves JL, Kamphuis C, Vernooij H, Araújo Jr JP, Grenfell RC, Juliano L and Dos Santos MV, 2020. Pathogen effects on milk yield and composition in chronic subclinical mastitis in dairy cows. The Veterinary Journal 262: 105473. https://doi.org/10.1016/j.tvjl.2020.105473
- Halasa T, Huijps K, Østerås O and Hogeveen H, 2007. Economic effects of bovine mastitis and mastitis management: A review. Veterinary Quarterly 29: 18-31. <u>https://doi.org/10.1080/0.1652176.2007.9695224</u>
- Hasan MS, Kober AK, Rana EA and Bari MS, 2022. Association of udder lesions with subclinical mastitis in dairy cows of Chattogram, Bangladesh. Advancements in Animal and Veterinary Sciences 10: 226-235. http://dx.doi.org/10.17582/journal.aavs/2022/10.2.226.235
- Hattermann K, Holzenburg E, Hans F, Lucius R, Held-Feindt J and Mentlein R, 2014. Effects of the chemokine CXCL12 and combined internalization of its receptors CXCR4 and CXCR7 in human MCF-7 breast cancer cells. Cell and Tissue Research 357: 253-266. <u>https://doi.org/10.1007/s00441-014-1823-y</u>
- He G, Ma M, Yang W, Wang H, Zhang Y and Gao MQ, 2017. SDF-1 in mammary fibroblasts of bovine with mastitis induces EMT and inflammatory response of epithelial cells. International Journal of Biological Sciences 13: 604. <u>https://doi.org/10.7150/ijbs.19591</u>
- Hofmeyr F and Oehme F, 1988. Textbook of Large Animal Surgery. The digestive system. Baltimore: Williams & Wilkins, pp: 448.
- Hogaboam CM, Steinhauser ML, Chensue SW and Kunkel SL, 1998. Novel roles for chemokines and fibroblasts in interstitial fibrosis. Kidney International 54: 2152-2159. <u>https://doi.org/10.1046/j.1523-1755.1998.00176.x</u>
- Houben EH, Dijkhuizen AA, Van Arendonk JA and Huirne RB, 1993. Short-and long-term production losses and repeatability of clinical mastitis in dairy cattle. Journal of Dairy Science 76: 2561-2578. <u>https://doi.org/10.3168/jds.S0022-0302(93)77591-8</u>
- Hussain R, MT Javed, A Khan and G Muhammad, 2013. Risks factors associated with sub-clinical mastitis in water buffaloes in Pakistan. Tropical Animal Health and Production 45: 1723-1729. http://dx.doi.org/10.1007/s11250-013-0421-4
- Ijaz M, Mehmood K, Durrani AZ, Sabir AJ, Abbas T and Ali S, 2014. Treatment of chronic mastitis in a dairy cow: A case report. Global Veterinaria 13: 01-04. <u>https://doi.org/10.5829/idosi.gv.2014.13.01.83150</u>

AGROBIOLOGICAL RECORDS ISSN: 2708-7182 (Print); ISSN: 2708-7190 (Online) Open Access Journal



- Jamali H, Barkema HW, Jacques M, Lavallée-Bourget EM, Malouin F, Saini V and Dufour S, 2018. Invited review: Incidence, risk factors, and effects of clinical mastitis recurrence in dairy cows. Journal of Dairy Science 101: 4729-4746. https://doi.org/10.3168/jds.2017-13730
- Ji Y, Xiao F, Zhu W, Liu SS, Feng X, Sun C, Lei L, Dong J, Khan A, Han W and Gu J, 2020. LysGH15 effectively control murine mastitis caused by *Staphylococcus aureus*. Pakistan Veterinary Journal 40: 519-522. http://dx.doi.org/10.29261/pakvetj/2020.056
- Kang H, Watkins G, Parr C, Douglas-Jones A, Mansel RE and Jiang WG, 2005. Stromal cell derived factor-1: its influence on invasiveness and migration of breast cancer cells in vitro, and its association with prognosis and survival in human breast cancer. Breast Cancer Research 7: 1-9. <u>https://doi.org/10.1186/bcr1022</u>
- Karthick K, 2020. Successful treatment of chronic fibrosed mastitis with treat fibrosis in cows by homeopathic remedies in: A review of 18 cases in field study. International Journal of Current Microbiology and Applied Sciences 9: 3194-3197. https://doi.org/10.20546/ijcmas.2020.908.364
- Khan A, R Hussain, MT Javed and F Mahmood, 2013. Molecular analysis of virulent genes (coa and spa) of Staphylococcus aureus involved in natural cases of bovine mastitis. Pakistan Journal of Agricultural Sciences 50: 739-743.
- Kobayashi K, Sato K, Kida T, Omori K, Hori M, Ozaki H and Murata T, 2014. Stromal cell–derived factor-1α/CXC chemokine receptor type 4 axis promotes endothelial cell barrier integrity via phosphoinositide 3-kinase and Rac1 activation. Arteriosclerosis, Thrombosis and Vascular Biology 34: 1716-1722. <u>https://doi.org/10.1161/ATVBAHA.114.303890</u>
- Kurjogi MM and Kaliwal BB, 2014. Epidemiology of bovine mastitis in cows of Dharwad district. International Scholarly Research Notices 2014: Article ID 968076. https://doi.org/10.1155/2014/968076
- Lancaster JE and Stuart P, 1949. Experiments on the transmission of Streptococcus agalactiae infection by milking with infected hands. Journal of Comparative Pathology and Therapeutics 59: 19. <u>https://doi.org/10.1016/S0368-1742(49)80002-6</u>
- Luker KE and Luker GD, 2006. Functions of CXCL12 and CXCR4 in breast cancer. Cancer Letters 238: 30-41. https://doi.org/10.1016/j.canlet.2005.06.021
- Makkar SS, 2017. A study of chronic teat fibrosis in bovines treated with homeopathy. Allgemeine Homöopathische Zeitung 262: 2-76. https://doi.org/10.1055/s-0037-1601149
- Miltenburg JD, De Lange D, Crauwels, APP, Bongers JH, Tielen MJM, Schukken YH and Elbers ARW, 1996. Incidence of clinical mastitis in a random sample of dairy herds in the southern Netherlands. Veterinary Record 139: 204-207. https://doi.org/10.1136/vr.139.9.204
- Mimoune N, Saidi R, Benadjel O, Khelef D and Kaidi R, 2021. Alternative treatment of bovine mastitis. Veterinarska Stanica 52: 639-49. https://doi.org/10.46419/vs.52.6.9
- Mohanty BK, Rath PK, Panda SK and Mishra BP, 2019. Pathomorphological studies of caprine mastitic udder. Journal of Entomology and Zoology Studies 7: 1208-1212.
- Mvrphy JM, 1947. The genesis of bovine udder infection and mastitis. ii. the occurrence of Streptococcal infection in a cow population during a seven-year period and its relationship to age. American Journal of Veterinary Research 8: 29-42.
- Nagahata H, Mukai T, Natsume Y, Okuda M, Ando T, Hisaeda K and Higuchi H, 2020. Effects of intramammary infusion of Bifidobacterium breve on mastitis pathogens and somatic cell response in quarters from dairy cows with chronic subclinical mastitis. Animal Science Journal 91: e13406. https://doi.org/10.1111/asj.13406
- Nieberle K and Cohrs P, 1966. Textbook of the Special Pathological Anatomy of Domestic Animals. Pergamon press Ltd., 1st Ed, London.
- Nyman AK, Ekman T, Emanuelson U, Gustafsson AH, Holtenius K, Waller KP and Sandgren CH, 2007. Risk factors associated with the incidence of veterinary-treated clinical mastitis in Swedish dairy herds with a high milk yield and a low prevalence of subclinical mastitis. Preventive Veterinary Medicine 78: 142-160. <u>https://doi.org/10.1016/j.prevetmed.2006.10.002</u>
- Oliveira SM, Ringshia RA, Legeros RZ, Clark E, Yost MJ, Terracio L and Teixeira CC, 2010. An improved collagen scaffold for skeletal regeneration. Journal of Biomedical Materials Research Part A 94: 371-379. <u>https://doi.org/10.1002/jbm.a.32694</u>
- Pal M, Regasa A and Gizaw F, 2019. Etiology, pathogenesis, risk factors, diagnosis and management of bovine mastitis: A comprehensive review. International Journal of Animal and Veterinary Sciences 6: 40-55.
- Plastige WN, Anderson EO and Weirether FJ, 1942. Infectious bovine mastitis. 8. The control of Streptococcus agalactia mastitis by a Segregation Program Based on Periodic Laboratory Tests. Storrs Agricultural Experiment Station 240.
- Plastridge WN, 1958. Bovine mastitis: a review. Journal of Dairy Science 41: 1141-1181. <u>https://doi.org/10.3168/jds.S0022-0302(58)91071-3</u>
- Qayyum A, JA Khan, R Hussain, A Khan, M Avais, N Ahmad and MF Hassan, 2016a. Molecular characterization of Staphylococcus aureus isolates recovered from natural cases of subclinical mastitis in Cholistani cattle and their antibacterial susceptibility. Pakistan Journal of Agricultural Sciences 53: 971-976.
- Qayyum A, JA Khan, R Hussain, M Avais, N Ahmed, A Khan and MS Khan, 2016b. Prevalence and association of possible risk factors with sub-clinical mastitis in cholistani cattle. Pakistan Journal of Zoology 48: 519-525.
- Radostits OM, Gay CC, Hinchcliff KW and Constable PD, 2007. A textbook of the diseases of cattle, horses, sheep, pigs and goats. Veterinary Medicine 10: 2045-2050.
- Rathod SU, Khodwe PM, Raibole RD and Vyavahare SH, 2009. Theloscopy-the advancement in teat surgery and diagnosis. Veterinary World 2: 34-37.
- Reza VH, Mehran FM, Majid MS and Hamid M, 2011. "Bacterial pathogens of intramammary infections in Azeri buffaloes of Iran and their antibiogram," African Journal of Agricultural Research 6: 2516–2521. <u>https://doi.org/10.5897/AJAR10.204</u>
- Riekerink RO, Barkema HW and Stryhn H, 2007. The effect of season on somatic cell count and the incidence of clinical mastitis. Journal of Dairy Science 90: 1704-1715. <u>https://doi.org/10.3168/jds.2006-567</u>

REVIEW ARTICLE



- Ruegg PL, 2017. A 100-Year review: Mastitis detection, management, and prevention. Journal of Dairy Science 100: 10381-10397. https://doi.org/10.3168/jds.2017-13023
- Rupp R and Boichard D, 2003. Genetics of resistance to mastitis in dairy cattle. Veterinary Research 34: 671-688. https://doi.org/10.1051/vetres:2003020
- Sarma O and Hussain J, 2021. Bovine Mastitis: An Overview. Vigyan Varta 2: 54-59.
- Schmenger A and Krömker V, 2020. Characterization, cure rates and associated risks of clinical mastitis in Northern Germany. Veterinary Sciences 7: 170. <u>https://doi.org/10.3390/vet_sci7040170</u>
- Schukken YH, Grommers FJ, Van de Geer D, Erb HN and Brand A, 1991. Risk factors for clinical mastitis in herds with a low bulk milk somatic cell count. 2. Risk factors for *Escherichia coli* and *Staphylococcus aureus*. Journal of Dairy Science 74: 826-832. https://doi.org/10.3168/jds.S0022-0302(91)78231-3
- Schukken YH, Günther J, Fitzpatrick J, Fontaine MC, Goetze L, Holst O and Pfizer, 2011. Mastitis Research Consortium. Hostresponse patterns of intramammary infections in dairy cows. Veterinary Immunology and Immunopathology 144: 270-289. https://doi.org/10.1016/j.vetimm.2011.08.022
- Semik-Gurgul E, Ząbek T, Kawecka-Grochocka, E, Zalewska M, Kościuczuk E and Bagnicka E, 2021. Epigenetic states of genes controlling immune responsiveness in bovine chronic mastitis. Annals of Animal Science. <u>https://doi.org/10.2478/aoas-2021-0061</u>
- Shah KA, Andrabi SA and Sumbul S, 2010. A study on homeopathic treatment of teat fibrosis in bovines. VetScan 5.
- Shah P, Shrivastava S, Gogoi P, Saxena S, Srivastava S, Singh RJ, Godara B, Kumar N and Gaur GK, 2022. Wasp venom peptide (Polybia MP-1) shows antimicrobial activity against multi drug resistant bacteria isolated from mastitic cow milk. International Journal of Peptide Research and Therapeutics 28: 44. <u>https://doi.org/10.1007/s10989-021-10355-0</u>
- Sladek Z, Rysanek D, Ryznarova H and Faldyna M, 2005. Neutrophil apoptosis during experimentally induced Staphylococcus aureus mastitis. Veterinary Research 36: 629–43.
- Sladek Z, Ryznarova H and Rysanek D, 2006. Macrophages of the bovine heifer mammary gland: morphological features during initiation and resolution of the inflammatory response. Anatomy Histology and Embryology 35: 116–24. https://doi.org/10.1111/j.1439-0264.2005.00647.x
- Sordillo LM and Streicher KL, 2002. Mammary gland immunity and mastitis susceptibility. Journal of Mammary Gland Biology and Neoplasia 7: 135-146.
- Suriyasathaporn W, Schukken YH, Nielen M and Brand A, 2000. Low somatic cell count: a risk factor for subsequent clinical mastitis in a dairy herd. Journal of Dairy Science 83: 1248-1255. <u>https://doi.org/10.3168/jds.S0022-0302(00)74991-5</u>
- Umadevi U, Saranya K, Madhu Mathi P and Umakanthan T, 2015. Successful treatment of fibrosed udder in cows. International Journal of Recent Scientific Research 6: 3574.
- Wang H, Yu G, Yu H, Gu M, Zhang J, Meng X and Li J, 2015. Characterization of TLR2, NOD2, and related cytokines in mammary glands infected by Staphylococcus aureus in a rat model. Acta Veterinaria Scandinavica 57: 1-6. <u>https://doi.org/10.1016/j.micpath.2020.104427</u>
- Wente N, Grieger AS, Klocke D, Paduch JH, Zhang Y, Leimbach S and Krömker V, 2020. Recurrent mastitis–persistent or new infections? Veterinary Microbiology 244: 108682. https://doi.org/10.1016/j.vetmic.2020.108682
- Xu J, Mora A, Shim H, Stecenko A, Brigham KL and Rojas M, 2007. Role of the SDF-1/CXCR4 axis in the pathogenesis of lung injury and fibrosis. American Journal of Respiratory Cell and Molecular Biology 37: 291-299. https://doi.org/10.1165/rcmb.2006-0187OC
- Youssef HA, 1999. Mastectomy as a radical treatment for some prevalent udder affections in goats in Al-Gasseem. Assiut Veterinary Medical Journal 41: 181-193.
- Zadoks RN, Allore HG, Barkema HW, Sampimon OC, Wellenberg GJ, Gröhn YT and Schukken YH, 2001. Cow-and quarterlevel risk factors for *Streptococcus uberis* and *Staphylococcus aureus* mastitis. Journal of Dairy Science 84: 2649-2663. https://doi.org/10.3168/jds.S0022-0302(01)74719-4
- Zenebe N, Habtamu T and Endale B, 2014. Study on bovine mastitis and associated risk factors in Adigrat, Northern Ethiopia. African Journal of Microbiology Research 8: 327–331. <u>https://doi.org/10.5897/AJMR2013.6483</u>
- Zou XZ, Liu T, Gong ZC, Hu CP and Zhang Z, 2017. MicroRNAs-mediated epithelial-mesenchymal transition in fibrotic diseases. European Journal of Pharmacology 796: 190-206. https://doi.org/10.1016/j.ejphar.2016.12.003